## Convenient Synthesis of $1\alpha,25$ -Dihydroxyvitamin $D_3$ from Vitamin $D_2^{1,2)}$

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(1S,6R)-1-Acetoxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclovitamin  $D_2$  (7) was synthesized from vitamin  $D_2$  by five steps. The new compound 7 was ozonized regioselectively and subsequently reduced, leading to (7E)-(1S,3S,5R,6R)-1-acetoxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclo-9,10-seco-23,24-dinor-7,10(19)-choladien-22-ol (11). The alcohol 11 obtained as a key intermediate was tosylated, iodinated, and coupled with 2-methyl-4-phenylsulfonyl-2-(tetrahydropyranyloxy)butane to give (1S,6R)-1-hydroxy-6-(1,3-benzodithiol-2-yloxy)-23-phenylsulfonyl-25-tetrahydropyranyloxy-3,5-cyclovitamin  $D_3$  (16). By desulfonylation and hydrolysis of the compound 16  $1\alpha,25$ -dihydoxyvitamin  $D_3$  was obtained selectively.

 $1\alpha, 25$ -Dihydroxyvitamin  $D_3$  (1a) has been widely used as a medicine of bone disease resulting from some metabolic disorder of vitamin D. Therefore 1a is an important compound, and a more convenient synthetic method has been desired (Chart 1). Since a total synthesis of 1a was reported by Semmler et al.,3) many reports<sup>4-10)</sup> about improved synthetic methods of **1a** have been published; such synthetic studies have continued. On the other hand, Abe et al. 11) reported that mouse myeloid leukemia cells could be induced to differentiate into macrophages by 1a. Since then a number of vitamin D derivatives have been synthesized, 12) for example aiming at the development of anticancer drugs. (13) So it is significant to find a practical synthesis of 1a, which will also be useful for synthesizing other vitamin D derivatives.

In the various synthetic methods of 1a, it is attractive to employ vitamin  $D_2$  as a starting material. Andrews et al.<sup>14)</sup> prepared 1a via regioselective ozonolysis of vitamin  $D_2$ -sulfur dioxide adducts<sup>15)</sup> (Scheme 1). In this method, since 5,6-trans vitamin D derivatives were obtained by thermal desulfonylation of the adducts, it was indispensable that the derivatives must be photoisomerized to convert to 5,6-cis vitamin D derivatives. Such information prompted us to study other methods via vitamin  $D_2$ .

In the previous work,<sup>2)</sup> we reported a practical method for synthesis of  $1\alpha$ -hydroxyvitamin  $D_3$  (**1b**) via 3,5-cyclovitamin  $D_3$  derivatives bearing a 1,3-benzodithiol-2-yl (BDT) group from vitamin  $D_3$ . By the application of the method, a new cyclovitamin  $D_2$  com-

pound was prepared. We discovered that the compound was subject to regioselective ozonolysis at the side-chain double bond. The key intermediate obtained was combined with a new side-chain moiety and cycloreversed, giving the target compound (1a) selectively. In our method, the process is not only short (11 steps), as is the Andrews' method described above, but it is easy to scale up, because 1a is readily obtained in high purity; in addition, the use of harmful sulfur dioxide and the procedure of photoisomerization are unnecessary.

On the other hand, a side-chain fragment was prepared in high yield by a modification of Tachibana's method. <sup>16)</sup> In this paper, further details are described.

Vitamin  $D_2$  (2) reacted with p-toluenesulfonyl chloride in dry pyridine to give vitamin D<sub>2</sub> tosylate (3) in 99% yield. The tosylate 3 was converted to (6R)-6hydroxy-3,5-cyclovitamin  $D_2$  (4) in 62% yield by hydrolyzing in aqueous acetone buffered with KHCO<sub>3</sub>.<sup>17)</sup> (6R)-6-(1,3-Benzodithiol-2-yloxy)-3,5-cyclovitamin  $D_2$ (5) was prepared from 4 and 1,3-benzodithiolylium tetrafluoroborate (BDTF)<sup>18)</sup> in pyridine-CH<sub>2</sub>Cl<sub>2</sub> (60% vield). The cyclovitamin 5 was oxidized with SeO<sub>2</sub>/ t-butyl hydroperoxide (TBHP) to give (1S,6R)-1-hydroxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclovitamin D<sub>2</sub>  $(6)^{19}$  and (6R)-1-oxo-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclovitamin D<sub>2</sub> in 59 and 11% yields, respectively. Compound 6 reacted with acetic anhydride in dry pyridine to give (1S,6R)-1-acetoxy-6-(1,3-benzodithiol-2yloxy)-3,5-cyclovitamin D<sub>2</sub> (7) in 97% yield (Scheme 2).

In the synthesis of  $1\alpha,25$ -dihydroxyvitamin  $D_3$  (1a) from vitamin  $D_2$  (2), regioselective double bond cleavage at its side chain part can be considered a key step. For that purpose, it is necessary to protect the conjugated triene system of vitamin D. There are protection methods leading vitamin D to cyclovitamin  $D^{7,20}$  and to sulfur dioxide adducts or other Diels–Alder adducts. In the former method, however, we feared that the regioselectivity in the ozonolysis of cyclovitamin D derivatives would be lowered owing to the presence of an *exo*-methylene group, which is liable to be oxidized at the A-ring part. In practice, (1S,6R)-1-acetoxy-6-methoxy-3,5-cyclovitamin  $D_2$  (8)<sup>21)</sup> was treated with ozone, followed by addition of dimethyl sulfide to

Scheme 1.

$$R_{2}O$$
 $R_{4}$ 
 $A: R_{3}=H, R_{4}=H$ 
 $2: R_{2}=H$ 
 $3: R_{2}=Ts$ 
 $6: R_{3}=BDT, R_{4}=OH$ 
 $7: R_{3}=BDT, R_{4}=OAc$ 
 $BDT = S$ 

give the aldehyde **9a** and the aldehyde **10a** in 24 and 42% yields, respectively. However, when compound **7** underwent ozonolysis in the same manner, the regioselectivity of the reaction was reversed to give the aldehyde **9b** and the aldehyde **10b** in 59 and 15% yields, respectively (Scheme 3).

Scheme 2.

We presume that the regionselectivity of the reaction is influenced by the differences in stable conformation between  $\mathbf{7}$  and  $\mathbf{8}$ . With (6R)-6-methoxy-3,5-cyclovitamin  $D_3$ , Sheves and Mazur<sup>20)</sup> reported that the methoxyl group is situated at one side of the exo-methylene group in its low energy conformation. In view of this, the oxidation of the exo-methylene group of  $\mathbf{7}$ , which has the much more bulky BDT group, (compared to the methoxyl group), can be assumed to be depressed.

The aldehyde **9b** was reduced with NaBH<sub>4</sub> to give the alcohol **11** in 88% yield. The latter was also obtained in 59% yield by the reductive decomposition of the ozonide of **7** with NaBH<sub>4</sub> in situ (Scheme 3). These **9b** and **11** compounds are useful as the intermediates of various vitamin D derivatives.

The alcohol 11 was tosylated in the manner described above to give the tosylate 12 in quantitative yield. The tosylate 12 reacted with NaI in refluxing acctone to afford the iodide 13 in quantitative yield (Scheme 3).

The preparation of 2-methyl-4-phenylsulfonyl-2-(tetrahydropyranyloxy)butane (15)<sup>16,22)</sup> as a side-chain fragment is shown in Scheme 4. Lithiation of methyl phenyl sulfone with butyllithium in THF, followed by the addition of 2,2-dimethyloxirane, gave 2-methyl-4-

phenylsulfonyl-2-butanol (14). The hydroxyl group of 14 was protected as a tetrahydropyranyl (THP) group in the usual manner to afford 15 in 87% yield from methyl phenyl sulfone.

TES= triethylsilyl

Lithiation of 15 with butyllithium in the presence of hexamethylphosphoric triamide (HMPA) in THF, followed by the addition of the iodide 13, gave 23-phenylsulfonyl derivative 16 simultaneously deacetylated. The reductive desulfonylation of 16 with sodium amalgam in methanol buffered with Na<sub>2</sub>HPO<sub>4</sub> provided cyclovitamin D<sub>3</sub> derivative 17 in 47% yield from 13. The compound 17 was hydrolyzed with 12-phosphomolybdic acid catalyst in aqueous dioxane and simultaneously subjected to the deprotection of the THP group, giving  $1\alpha$ ,25-dihydroxyvitamin D<sub>3</sub> (1a) in 30% yield (Scheme 5). In the <sup>1</sup>H NMR (400 MHz) spectra of 1a, the signals attributed to the 5,6-trans isomer were not detected.

## Experimental

<sup>1</sup>H NMR spectra were recorded on a Bruker AMX-400 spectrometer in CDCl<sub>3</sub> with TMS as an internal standard. IR spectra were measured on a Perkin–Elmer 1720X spectrometer. Mass spectra were obtained at 20 eV on a Hitachi M-80 spectrometer. Fuji Silysia Chemical BW-820H (70—200 mesh) was used for SiO<sub>2</sub> column chromatography. Melting points are uncorrected.

Vitamin D<sub>2</sub> Tosylate (3). Compound 3 was synthesized according to the literature: <sup>17)</sup> Mp 89.5—90 °C (decomp); <sup>1</sup>H NMR  $\delta$ =0.54 (3H, s, 18-H<sub>3</sub>), 0.82 and 0.84 (each 3H, d, J=6.5 Hz, 26-H<sub>3</sub> and 27-H<sub>3</sub>), 0.91 (3H, d, J=6.8 Hz, 28-H<sub>3</sub>), 1.01 (3H, d, J=6.6 Hz, 21-H<sub>3</sub>), 2.45 (3H, s, CH<sub>3</sub>(tosyl)), 4.70 (1H, m, 3-H), 4.81 (1H, m(sharp), 19-H), 5.02 (1H, m(sharp), 19-H), 5.19 (2H, m, 22-H and 23-H), 5.96 (1H, d, J=11.2 Hz, 7-H), 6.09 (1H, d, J=11.2 Hz, 6-H), 7.33 (2H, m, Ar-H<sub>2</sub>), and 7.80 (2H, m, Ar-H<sub>2</sub>).

(6R)-6-Hydroxy-3,5-cyclovitamin  $D_2$  (4). Compound 4 was synthesized according to the literature: <sup>17)</sup> IR (neat) 3392, 1650, 1457, 1372, 1052, 1024, 999, 973, and 869 cm<sup>-1</sup>; <sup>1</sup>H NMR δ=0.55 (3H, s, 18-H<sub>3</sub>), 0.66 (1H, m, 4-H), 0.82 and 0.84 (each 3H, d, J=6.5 Hz, 26-H<sub>3</sub> and 27-H<sub>3</sub>), 0.92 (3H, d, J=6.8 Hz, 28-H<sub>3</sub>), 0.90—0.94 (1H, m, 4-H, overlapped with 28-H<sub>3</sub>), 1.02 (3H, d, J=6.6 Hz, 21-H<sub>3</sub>), 4.93 (3H, m, 6-H, 7-H, and 19-H), 4.99 (1H, m(sharp), 19-H), and 5.19 (2H, m, 22-H and 23-H).

(6R)-6-(1,3-Benzodithiol-2-yloxy)-3,5-cyclovitamin  $D_2$  (5). To a solution of 4 (1.25 g, 3.15 mmol) in dry  $CH_2Cl_2$  (28 ml) was added dry pyridine (2.5 ml).

Scheme 3.

The solution was cooled at -10 °C and BDTF (1.54 g, 6.43 mmol) was added. After stirring for 8 h at -5—0 °C, triethylamine (9 ml) was added. The solution was stirred overnight at room temperature and concentrated in vacuo. The residue was extracted with hexane, washed with H<sub>2</sub>O and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was washed with acetone to give 5 (1.04 g, 60%): Mp 126.5—127 °C (from acetone); IR (KBr) 3060, 3025, 1669, 1646, 1461, 1446, 1371, 1037, 1020, 970, 947, 884, and 740 cm<sup>-1</sup>; <sup>1</sup>H NMR  $\delta$ =0.52 (3H, s, 18-H<sub>3</sub>), 0.79 (1H, m, 4-H), 0.82 and 0.84 (each 3H, d, J=6.5 Hz,  $26-H_3$  and  $27-H_3$ ),  $0.92 (3H, d, J=6.8 Hz, 28-H_3), 0.93 (1H, m, 4-H), 1.01 (3H, m, 4-H)$ d, J=6.6 Hz, 21-H<sub>3</sub>), 4.59 (1H, d, J=9.5 Hz, 6-H), 4.83 (1H, m(sharp), 19-H), 5.09 (1H, m(sharp), 19-H), 5.15 (1H, d, J=9.5 Hz, 7-H), 5.19 (2H, m, 22-H and 23-H), 6.55 (1H, s, SCHS), 7.06 (2H, m, Ar-H<sub>2</sub>), and 7.30 (2H, m, Ar-H<sub>2</sub>). Found: m/z 548.3126. Calcd for  $C_{35}H_{48}OS_2$ : M, 548.3144.

(1S,6R)-1-Hydroxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclovitamin  $D_2$  (6). A mixture of 70% TBHP-H<sub>2</sub>O (0.536 g, 4.16 mmol) and CH<sub>2</sub>Cl<sub>2</sub> (6 ml) was dried (MgSO<sub>4</sub>). The drying agent was filtered off and washed with CH<sub>2</sub>Cl<sub>2</sub> (6 ml). To the combined CH<sub>2</sub>Cl<sub>2</sub> solutions was added SeO<sub>2</sub> (0.115 g, 1.04 mmol), and after stirring for 15 min, a solution of 5 (1.15 g, 2.10 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (12 ml) was added to the mixture at 20 °C. The mixture was stirred for ca. 15 min at 20-25 °C and 5% aqueous NaOH (10 ml) was added. After vigorous stirring for 10 min, the CH<sub>2</sub>Cl<sub>2</sub> phase was washed with H<sub>2</sub>O and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub> (31 g), eluting with hexane-EtOAc (30:1—10:1) to give 6 (0.70 g, 59%): Mp 142—143 °C; IR (KBr) 3540, 1656, 1445, 1370, 1078, 1035, 752, and 718 cm<sup>-1</sup>; <sup>1</sup>H NMR  $\delta$ =0.53 (3H, s, 18-H<sub>3</sub>), 0.64 (1H, m, 4-H), 0.82 and 0.84 (each 3H, d, J=6.5 Hz, 26-H<sub>3</sub> and 27- $H_3$ ), 0.92 (4H, d (J=6.8 Hz) and m, 28- $H_3$  and 4-H), 1.02 (3H, d, J=6.6 Hz, 21-H<sub>3</sub>), 4.16 (1H, m, 1-H), 4.66 (1H, d, d)

 $J\!=\!9.6$  Hz, 6-H), 5.05 (1H, d,  $J\!=\!9.6$  Hz, 7-H), 5.11 (1H, d,  $J\!=\!2$  Hz, 19-H), 5.20 (2H, m, 22-H and 23-H), 5.27 (1H, d,  $J\!=\!2$  Hz, 19-H), 6.50 (1H, s, SCHS), 7.08 (2H, m, Ar-H<sub>2</sub>), and 7.31 (2H, m, Ar-H<sub>2</sub>). Found: m/z 564.3106. Calcd for  $\rm C_{35}H_{48}O_2S_2$ : M, 564.3093.

At this time 0.13 g (11%) of (6R)-1-oxo-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclovitamin D<sub>2</sub> was given as a less polar by-product:  $^1\mathrm{H}$  NMR  $\delta$ =0.48 (3H, s, 18-H<sub>3</sub>), 0.53 (1H, m, 4-H), 0.82 and 0.83 (each 3H, d, J=6.6 Hz, 26-H<sub>3</sub> and 27-H<sub>3</sub>), 0.92 (3H, d, J=6.8 Hz, 28-H<sub>3</sub>), 1.01 (3H, d, J=6.6 Hz, 21-H<sub>3</sub>), 4.64 (1H, d, J=9.6 Hz, 6-H), 5.04 (1H, d, J=9.6 Hz, 7-H), 5.19 (2H, m, 22-H and 23-H), 5.59 (1H, m(sharp), 19-H), 5.97 (1H, m(sharp), 19-H), 6.48 (1H, s, SCHS), 7.09 (2H, m, Ar-H<sub>2</sub>), 7.30 (1H, m, Ar-H), and 7.34 (1H, m, Ar-H). Found: m/z 562.2931. Calcd for C<sub>35</sub>H<sub>46</sub>O<sub>2</sub>S<sub>2</sub>: M, 562.2936.

(1S, 6R)-1-Acetoxy-6-(1, 3-benzodithiol-2-vloxy)-3,5-cyclovitamin  $D_2$  (7). To a solution of 6 (3.78) g, 6.69 mmol) in dry pyridine (38 ml) was added acetic anhydride (3.42 g, 33.5 mmol), and the mixture was stirred overnight at room temperature. Crushed ice was added, and after stirring for 1 h, the mixture was extracted with ether. The ether solution was washed with aqueous CuSO<sub>4</sub>, H<sub>2</sub>O, and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub> (120 g), eluting with hexane–EtOAc (30:1) to give 7 (3.95 g, 97%): <sup>1</sup>H NMR  $\delta = 0.53$  (3H, s, 18-H<sub>3</sub>), 0.71 (1H, m, 4-H), 0.82 and 0.84 (each 3H, d, J=6.5 Hz,  $26-H_3$  and  $27-H_3$ ), 0.92 (3H, d,  $J=6.8 \text{ Hz}, 28-\text{H}_3$ , 0.97 (1H, m, 4-H), 1.02 (3H, d, J=6.6Hz, 21-H<sub>3</sub>), 2.07 (3H, s, OCOCH<sub>3</sub>), 4.65 (1H, d, J=9.8 Hz, 6-H), 4.93 (1H, d, J=2 Hz, 19-H), 5.08 (1H, d, J=9.8 Hz, 7-H), 5.20 (2H, m, 22-H and 23-H), 5.28 (1H, d, J=2 Hz, 19-H), 6.51 (1H, s, SCHS), 7.08 (2H, m, Ar-H<sub>2</sub>), and 7.31 (2H, m, Ar-H<sub>2</sub>). Found: m/z 606.3195. Calcd for  $C_{37}H_{50}O_3S_2$ : M, 606.3197.

Ozonolysis of (1S,6R)-1-Acetoxy-6-methoxy-3,5-cyclovitamin  $D_2$  (8). Compound 8 was prepared according to the procedure of Ref.  $21.^{23}$  Ozone was bubbled into a mixture of 8  $(0.28~{\rm g},~0.60~{\rm mmol})$ , dry pyridine  $(0.1~{\rm ml})$ , and dry CH<sub>2</sub>Cl<sub>2</sub>  $(32~{\rm ml})$  at  $-70~{\rm ^{\circ}C}$ . The reaction was monitored by TLC (hexane–EtOAc, 5:1). After purging the system with N<sub>2</sub>, dimethyl sulfide  $(0.5~{\rm ml})$  was added. The solution was warmed to room temperature, washed with aqueous CuSO<sub>4</sub> and brine, dried (MgSO<sub>4</sub>), and concentrated

BDTO...

BDTO...

BDTO...

$$R_7$$

OH

 $R_7$ 

1a

 $R_7$ 

15

 $R_7$ 

17:  $R_7$ =H

Scheme 5.

in vacuo. The residue was chromatographed on SiO<sub>2</sub> (27 g), eluting with hexane–EtOAc (7:1—5:1) to give 57 mg (24%) of (7E)-(1S,3S,5R,6R)-1-acetoxy-6-methoxy-3,5-cyclo-9,10-seco-23,24-dinor-7,10(19)-choladien-22-al ( $\bf 9a$ ) as the less polar component and 100 mg (42%) of (7E)-(1S,3S,5R,6R)-1-acetoxy-6-methoxy-3,5-cyclo-10-oxo-9,10-seco-19,23,24-trinorchol-7-en-22-al ( $\bf 10a$ ) as the more polar component.

Aldehyde **9a**: <sup>1</sup>H NMR  $\delta$ =0.59 (3H, s, 18-H<sub>3</sub>), 0.70 (1H, m, 4-H), 0.97 (1H, m, 4-H), 1.14 (3H, d, J=6.9 Hz, 21-H<sub>3</sub>), 2.09 (3H, s, OCOCH<sub>3</sub>), 3.26 (3H, s, OCH<sub>3</sub>), 4.16 (1H, d, J=9.2 Hz, 6-H), 4.98 (1H, d, J=1.6 Hz, 19-H), 5.03 (1H, d, J=9.2 Hz, 7-H), 5.22 (1H, m, 1-H), 5.24 (1H, d, J=1.6 Hz, 19-H, overlapped with 1-H), and 9.60 (1H, d, J=3.2 Hz, CHO). Found: m/z 400.2615. Calcd for C<sub>25</sub>H<sub>36</sub>O<sub>4</sub>: M, 400.2611.

Aldehyde **10a**:  $^{1}$ H NMR  $\delta$ =0.57 (3H, s, 18-H<sub>3</sub>), 1.14 (3H, d, J=6.9 Hz, 21-H<sub>3</sub>), 1.16 (1H, m, 4-H, overlapped with 21-H<sub>3</sub>), 2.13 (3H, s, OCOCH<sub>3</sub>), 3.19 (3H, s, OCH<sub>3</sub>), 4.63 and 4.67 (2H, ABq, J=9.5 Hz, 6-H and 7-H), 5.18 (1H, m, 1-H), and 9.59 (1H, d, J=3.2 Hz, CHO). Found: m/z 402.2422. Calcd for C<sub>24</sub>H<sub>34</sub>O<sub>5</sub>: M, 402.2404.

Ozonolysis of (1S,6R)-1-Acetoxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclovitamin  $D_2$  (7). Compound 7 (0.24 g, 0.40 mmol) was treated with ozone as described above to give 126 mg (59%) of (7E)-(1S,3S,5R,6R)-1-acetoxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclo-9,10-seco-23,24-dinor-7,10(19)-choladien-22-al (9b) and 31 mg (15%) of (7E)-(1S,3S,5R,6R)-1-acetoxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclo-10-oxo-9,10-seco-19,23,24-trinorchol-7-en-22-al (10b).

Aldehyde **9b**: <sup>1</sup>H NMR  $\delta$ =0.56 (3H, s, 18-H<sub>3</sub>), 0.72 (1H, m, 4-H), 0.97 (1H, m, 4-H), 1.14 (3H, d, J=6.9 Hz, 21-H<sub>3</sub>), 2.07 (3H, s, OCOCH<sub>3</sub>), 4.64 (1H, d, J=9.5 Hz, 6-H), 4.94 (1H, d, J=2 Hz, 19-H), 5.13 (1H, d, J=9.5 Hz, 7-H), 5.17 (1H, m, 1-H), 5.27 (1H, d, J=2 Hz, 19-H), 6.52 (1H, s, SCHS), 7.08 (2H, m, Ar-H<sub>2</sub>), 7.31 (2H, m, Ar-H<sub>2</sub>), and 9.59 (1H, d, J=3.1 Hz, CHO). Found: m/z 538.2203. Calcd for C<sub>31</sub>H<sub>38</sub>O<sub>4</sub>S<sub>2</sub>: M, 538.2208.

Aldehyde **10b**: <sup>1</sup>H NMR  $\delta$  = 0.56 (3H, s, 18-H<sub>3</sub>), 1.13 (3H, d, J = 6.9 Hz, 21-H<sub>3</sub>), 1.17 (1H, m, 4-H), 2.11 (3H, s, OCOCH<sub>3</sub>), 4.78 and 5.09 (2H, ABq, J = 9.6 Hz, 6-H and 7-H), 5.14 (1H, m, 1-H), 6.42 (1H, s, SCHS), 7.09 (2H, m, Ar-H<sub>2</sub>), 7.30 (2H, m, Ar-H<sub>2</sub>), and 9.59 (1H, d, J = 3.2 Hz, CHO). Found: C, 66.44; H, 6.84; S, 11.38%. Calcd for C<sub>30</sub>H<sub>36</sub>O<sub>5</sub>S<sub>2</sub>: C, 66.64; H, 6.71; S, 11.86%.

 $\begin{array}{lll} (7E)\text{-}(1S,\!3S,\!5R,\!6R)\text{-}1\text{-}Acetoxy\text{-}6\text{-}(1,\!3\text{-}benzodithi-}\\ \text{ol-2-yloxy})\text{-}3,\!5\text{-}cyclo\text{-}9,\!10\text{-}seco\text{-}23,\!24\text{-}dinor\text{-}7,\!10(19)\text{-}\\ \text{choladien-}22\text{-}ol\ (11). & Method\ A: & To\ a\ solution\\ \text{of}\ 9b\ (100\ \text{mg},\ 0.19\ \text{mmol})\ \text{in}\ CH_2Cl_2\text{-}EtOH\ (1:1,\ 5\ \text{ml})\\ \text{was}\ added\ NaBH_4\ (21\ \text{mg},\ 0.56\ \text{mmol}). & The\ \text{mixture}\ \text{was} \end{array}$ 

stirred for 1 h at room temperature and concentrated in vacuo. Ether and cold 1 moldm<sup>-3</sup> HCl was successively added to the residue, and the mixture was stirred for 20 min. The ether solution was washed with aqueous NaHCO<sub>3</sub> and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub> (9.5 g), eluting hexane-EtOAc (7:3) to give 11 (88 mg, 88%): Mp 159-160 °C; IR (KBr) 3535, 3054, 3027, 1719, 1665, 1445, 1370, 1256, 1045, 1026, 997, 968, and 759 cm<sup>-1</sup>; <sup>1</sup>H NMR  $\delta$ =0.54 (3H, s, 18-H<sub>3</sub>), 0.71 (1H, m, 4-H), 0.97 (1H, m, 4-H), 1.06 (3H, d, J=6.6 Hz, 21-H<sub>3</sub>), 2.07 (3H, s, OCOCH<sub>3</sub>), 3.40 and 3.65 (2H, each m, 22-H<sub>2</sub>), 4.66 (1H, d, J=9.6 Hz, 6-H), 4.94 (1H, d)d, J=2 Hz, 19-H), 5.09 (1H, d, J=9.6 Hz, 7-H), 5.17 (1H, m, 1-H), 5.27 (1H, d, J=2 Hz, 19-H), 6.51 (1H, s, SCHS), 7.08 (2H, m, Ar-H<sub>2</sub>), and 7.31 (2H, m, Ar-H<sub>2</sub>). Found: C, 68.64; H, 7.64; S, 12.08%. Calcd for C<sub>31</sub>H<sub>40</sub>O<sub>4</sub>S<sub>2</sub>: C, 68.85; H, 7.46; S, 11.86%.

Method B: Compound 7 (1.00 g, 1.65 mmol) in 1% pyridine—CH<sub>2</sub>Cl<sub>2</sub> (30 ml) was treated with ozone at -70 °C, followed by bubbling of N<sub>2</sub> and addition of NaBH<sub>4</sub> (200 mg, 5.29 mmol) in EtOH (7.5 ml). The solution was warmed to room temperature and poured into cold 1 mol dm<sup>-3</sup> HCl. After stirring for 30 min, the CH<sub>2</sub>Cl<sub>2</sub> layer was separated and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic extracts were washed with aqueous NaHCO<sub>3</sub> and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub> (50 g), eluting with hexane—EtOAc (7:2) to give 11 (0.53 g, 59%). The spectral data were consistent with those obtained by method A.

(7E)-(1S,3S,5R,6R)-1-Acetoxy-6-(1,3-benzodithiol-2-yloxy)-3,5-cyclo-9,10-seco-23,24-dinor-7,10(19)choladien-22-yl Tosylate (12). To a solution of 11 (0.26 g, 0.48 mmol) in dry pyridine (5 ml) was added p-TsCl (0.45 g, 2.4 mmol) at 5 °C, and the mixture was stirred overnight at 0—5 °C. Crushed ice was added, and after stirring for 1 h, the mixture was extracted with ether. The ether extract was washed with aqueous CuSO<sub>4</sub> and brine, bried (MgSO<sub>4</sub>), and concentrated in vacuo to give 12 (0.33 g, quant):  ${}^{1}\text{H NMR}$  $\delta\!=\!0.48$  (3H, s, 18-H<sub>3</sub>), 0.70 (1H, m, 4-H), 0.95 (1H, m, 4-H), 1.00 (3H, d, J = 6.6 Hz, 21-H<sub>3</sub>), 2.06 (3H, s, OCOCH<sub>3</sub>), 2.45 (3H, s, CH<sub>3</sub> (tosyl)), 3.81 and 3.98 (2H, each m, 22-H<sub>2</sub>), 4.63 (1H, d, J=9.5 Hz, 6-H), 4.93 (1H, d, J=2 Hz, 19-H), 5.08 (1H, d, J=9.5 Hz, 7-H), 5.16 (1H, m, 1-H), 5.25 (1H, m, 1-H)d, J=2 Hz, 19-H), 6.50 (1H, s, SCHS), 7.08 (2H, m, Ar-H<sub>2</sub>), 7.30 (2H, m, Ar-H<sub>2</sub>), 7.35 (2H, m, Ar-H<sub>2</sub>(tosyl)), and 7.79 (2H, m, Ar-H<sub>2</sub>(tosyl)). Found: C, 65.61; H, 6.71; S, 13.67%. Calcd for  $C_{38}H_{46}O_6S_3$ : C, 65.68; H, 6.67; S, 13.84%.

(7E)-(1S,3S,5R,6R)-1-Acetoxy-6-(1,3-benzodithiol-2-yloxy)-22-iodo-3,5-cyclo-9,10-seco-23,24-dinor-

A solution of **12** (0.30 g, 7,10(19)-choladiene (13). 0.43 mmol) and NaI (0.32 g, 2.1 mmol) in dry acetone (9 ml) was refluxed with stirring for 5 h. After being cooled. the solution was concentrated in vacuo and the residue was extracted with ether. The ether extract was washed with aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo to give 13 (0.28 g, quant):  ${}^{1}\text{H NMR }\delta = 0.56$ (3H, s, 18-H<sub>3</sub>), 0.71 (1H, m, 4-H), 0.97 (1H, m, 4-H), 1.04  $(3H, d, J=6.3 Hz, 21-H_3), 2.07 (3H, s, OCOCH_3), 3.18 and$ 3.33 (2H, each m, 22-H<sub>2</sub>), 4.64 (1H, d, J=9.5 Hz, 6-H), 4.94 (1H, d, J=2 Hz, 19-H), 5.10 (1H, d, J=9.5 Hz, 7-H), 5.17 (1H, m, 1-H), 5.27 (1H, d, J=2 Hz, 19-H), 6.51 (1H, s, SCHS), 7.08 (2H, m, Ar-H<sub>2</sub>), and 7.31 (2H, m, Ar-H<sub>2</sub>). Found: m/z 650.1368. Calcd for  $C_{31}H_{39}O_3IS_2$ : M, 650.1383.

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2-Methyl-4-phenylsulfonyl-2-(tetrahydropyranyloxy)butane (15). To a solution of methyl phenyl sulfone (1.00 g, 6.40 mmol) in dry THF (20 ml) was added 1.6 mol dm<sup>-3</sup> butyllithium-hexane (6.0 ml, 9.6 mmol) at -25—-20 °C under N<sub>2</sub>. After stirring for 20 min, 2,2-dimethyloxirane (1.16 g, 16.1 mmol) was added at -20 °C, and the mixture was stirred for 2 h at 2 °C and quenched by addition of a sat. NH<sub>4</sub>Cl solution. The organic layer was separated and the aqueous layer was extracted with EtOAc. The combined organic layers were washed with brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was crude 14, which was used without further purification. An analytical sample was purified by SiO<sub>2</sub> column chromatography (CHCl<sub>3</sub>-EtOAc, 10:1), and the spectral data were identical with those reported.<sup>22)</sup> According to the literature<sup>16)</sup> the crude **14** (1.51 g) was converted to **15** (1.74 g) in 87% yield from methyl phenyl sulfone: Mp 85—86 °C (lit, 22) mp 84-85 °C).

(1S,6R)-1-Hydroxy-6-(1,3-benzodithiol-2-yloxy)-23-phenylsulfonyl-25-tetrahydropyranyloxy-3,5-cyclovitamin D<sub>3</sub> (16). 1.6 mol dm<sup>-3</sup> Butyllithium-hexane (3.4 ml, 5.4 mmol) was added to a mixture of 15 (1.73 g. 5.54 mmol), dry HMPA (0.4 ml), and dry THF (8.6 ml) at -50 °C under N<sub>2</sub>. After stirring for 20 min at -25—-20 °C, a solution of **13** (0.74 g, 1.14 mmol) in dry THF (3.6 ml) was added, while the temperature was held constant, and the mixture was stirred for 2 h at -20 °C and quenched by addition of a sat. NH<sub>4</sub>Cl solution. The organic phase was washed with brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub> (87 g), eluting with hexane-EtOAc (5:2) to give crude 16 (0.78 g) as a foam, which was used without further purification. The <sup>1</sup>H NMR spectra indicated that the foam was contaminated with small amounts of impurities: <sup>1</sup>H NMR  $\delta = 0.46$ , 0.47, 0.48, and 0.50 (12H, each s,  $(18-H_3)\times 4$ ), 4.16 (4H, m,  $(1-H)\times 4$ , 4.68 (8H, m, (6-H, and CH(THP))×4), 5.08 (8H, m,  $(7-H \text{ and } 19-H)\times 4)$ ,  $5.26 (4H, m(sharp), (19-H)\times 4)$ , 6.48 (4H, s, (SCHS) $\times$ 4), 7.07 (8H, m, (Ar-H<sub>2</sub>) $\times$ 4), 7.31  $(8H, m, (Ar-H_2)\times 4), 7.55 (8H, m, (Ar-H_2(SO_2Ph))\times 4),$ 7.62 (4H, m, (Ar-H(SO<sub>2</sub>Ph))×4), and 7.89 (8H, m, (Ar- $H_2(SO_2Ph))\times 4).$ 

(1S,6R)-1-Hydroxy-6-(1,3-benzodithiol-2-yloxy)-25-tetrahydropyranyloxy-3,5-cyclovitamin  $D_3$  (17). To a solution of crude 16 (0.78 g) in THF-MeOH (0.8 ml. 27 ml respectively) was successively added Na<sub>2</sub>HPO<sub>4</sub> (1.40 g, 9.86 mmol) and 5% Na-Hg (4.52 g, 9.83 mmol), and the mixture was stirred for 3 h at room temperature. The supernatant was concentrated in vacuo and extracted with ether. The ether solution was washed with H<sub>2</sub>O and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub> (28 g), eluting with hexane-EtOAc (5:1) to afford **17** (0.35 g, 47% from **13**): <sup>1</sup>H NMR  $\delta = 0.54$  (3H, s, 18-H<sub>3</sub>), 0.64 (1H, m, 4-H), 0.92 (3H, d, J = 6.3Hz, 21-H<sub>3</sub>), 0.90—0.95 (1H, m, 4-H, overlapped with 21-H<sub>3</sub>), 1.19 and 1.21 (6H, each s, 26-H<sub>3</sub> and 27-H<sub>3</sub>), 3.44 and 3.95 (2H, each m, CH<sub>2</sub>(THP)), 4.16 (1H, m, 1-H), 4.67 (1H, d, J=9.6 Hz, 6-H), 4.72 (1H, m, CH(THP)), 5.06 (1H, d, J=9.6Hz, 7-H), 5.11 (1H, d, J=2 Hz, 19-H), 5.27 (1H, d, J=2 Hz, 19-H), 6.50 (1H, s, SCHS), 7.08 (2H, m, Ar-H<sub>2</sub>), and 7.31 (2H, m, Ar-H<sub>2</sub>). Found: C, 71.75; H, 8.91; S, 9.75%. Calcd for C<sub>39</sub>H<sub>56</sub>O<sub>4</sub>S<sub>2</sub>: C, 71.74; H, 8.64; S, 9.82%.

 $1\alpha,25$ -Dihydroxyvitamin D<sub>3</sub> (1a). To a solution of 17 (0.55 g, 0.84 mmol) in 5% H<sub>2</sub>O-dioxane (16.5 ml) was added phosphomolybdic acid (53.6 mg, 0.028 mmol as a H<sub>3</sub>PMo<sub>12</sub>O<sub>40</sub>•5H<sub>2</sub>O), and the mixture was stirred for 3 h at 30 °C. After being cooled to 15 °C, the mixture was poured into a sat. NaHCO3 solution and extracted with EtOAc. The EtOAc solution was washed with H<sub>2</sub>O and brine, dried (MgSO<sub>4</sub>), and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub>, eluting CHCl<sub>3</sub>-MeOH (30:1) to give 1a (104 mg, 30%). The <sup>1</sup>H NMR spectra were in accord with those reported:<sup>24)</sup> <sup>1</sup>H NMR  $\delta$ =0.55 (3H, s), 0.94 (3H, d, J=6.4 Hz), 1.21 (6H, s), 2.31 (1H, dd, J=13.4 and 6.5 Hz), 2.60 (1H, dd, J=13.4 and 3.2 Hz), 2.82 (1H, dd, J=12.0 and3.7 Hz), 4.23 (1H, m), 4.43 (1H, m), 5.00 (1H, m(sharp)), 5.33 (1H, m(sharp)), 6.02 (1H, d, J=11.2 Hz), and 6.38 (1H, d, J=11.2 Hz); mp 115—117 °C (from HCO<sub>2</sub>Me) (lit, <sup>25)</sup> mp 117—118 °C (from HCO<sub>2</sub>Me));  $[\alpha]_D^{20} + 45.0^\circ$  (c 0.5, EtOH)  $(\text{lit},^{8}) [\alpha]_{D}^{25} + 47.9^{\circ} (c \ 0.5, \text{ EtOH})).$ 

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- 19) Compound **6** (163 mg) was hydrolyzed similarly to compound **17** and purified by preparative layer chromatography (CHCl<sub>3</sub>–MeOH, 15:1), giving  $1\alpha$ -hydroxyvitamin D<sub>2</sub> (53 mg, 45%). The <sup>1</sup>H NMR spectra were in accord with that reported:<sup>7) 1</sup>H NMR  $\delta$ =0.56 (3H, s), 0.82 and 0.84 (each 3H, d, J=6.5 Hz), 0.92 (3H, d, J=6.8 Hz), 1.02 (3H, d, J=6.6 Hz), 4.23 (1H, m), 4.43 (1H, m), 5.00 (1H, m (sharp)), 5.20 (2H, m), 5.32 (1H, m (sharp)), 6.01 (1H, d, J=11.3 Hz), and 6.38 (1H, d, J=11.3 Hz); mp 141.5—142.5 °C (from acetone) (mp 141—143 °C, see: D. R. Andrews, D. H. R.

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